# Castlemaine Naturalist

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Monthly newsletter of the Castlemaine Field Naturalists Club Inc.



The critically endangered Corroboree Frog photo by Noel Young

# July Meeting Report: Speaker, Dr Matt West "Protecting frogs from the Chytrid fungus"

At our July monthly meeting, we were privileged to have a talk by amphibian ecologist, Dr Matt West (BSc, M Reprod Sci, PhD), who provided us with an update on the status of research into the chytrid fungus and its impact on frog populations as well as strategies for protecting frogs from this deadly fungus. Matt is a wildlife consultant for "Wild Research" with projects aimed at protecting threatened species with partnerships including the Threatened Species Recovery Hub, Zoos Victoria, DEECA, CMAs and traditional owners. He is also a Research Fellow at the University of Melbourne School of Biosciences.

Matt first gave us some background history. It was when scientists gathered at the First World Congress of Herpetology in 1989, that it became obvious that there was a serious decline in frog populations worldwide. Habitat change was soon identified as a major cause of amphibian decline, but it was the finding of a large number of dead frogs in the rivers of the Sierra Nevada that pointed to disease as a cause and the chytrid fungus was identified as the pathogen. Matt showed us images of the fungus and explained its life cycle. Water-borne fungal spores attach and burrow into the frog's skin, where they mature and develop into a sporangium which after a few days releases another batch of spores into the water. Infection with this fungus causes the frogs to shed layers of skin, disrupting normal skin function of gas exchange and osmoregulation. The effect on ion transport leads to cardiac arrest and death. An obvious sign of this disease, called Chytridiomycosis, is the frog's lack of ability to right itself after being turned over.

Detection of the chytrid fungus can now be made from a swab of the frog using a genetic lab test. A map showing sites of chytrid detection revealed

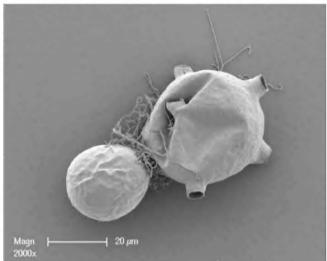


Dr Matt West collecting samples at one of his study locations. Photo by Michael Williams

the enormous scale of the problem globally. Chytrid has been shown to infect over 700 amphibian species, many in decline and some likely heading for extinction. In Australia, chytrid was first detected in Brisbane in 1978 and has now spread along the east and south-east regions including Tasmania as well as in southern WA.

An important finding was that not all frog species are susceptible to Chytridiomycosis despite being infected. So many researchers including Matt, are studying the factors that influence chytrid disease outcome. Environmentally, temperature and salinity have been found to be important,

with temperatures over 28C causing death of the fungus. High salinity, acidic or basic conditions also reduce chytrid disease. frogs themselves have natural defences from resident skin bacteria, microbial peptides and complement proteins that can cause inflammation and lyse the pathogen. Some frog species have robust immune defences. currently studying the range of genetic variations in the chytrid fungus. These have been shown to from which new spores are released. affect the number of spores released ie the virulence of the



Chytrid fungus sporangium, showing tubules

Photo: Matt West

fungus. So Matt explained that there is a disease triangle between pathogen virulence, host susceptibility and environmental factors.

Matt then explained how we can use this knowledge to design strategies for protecting frogs from chytrid disease. The fungus is obviously now too widespread for tackling eradication or containment (sound familiar?) but control and mitigation are possible. Individual frogs can be treated with antifungals, heat and salt but this is not possible at a population level. The fungus can survive in wet conditions for up to 3 months and there are multiple host frog species, so a cleaned frog returned to the wild is likely to be rapidly reinfected. Matt described projects for three species of frogs that are in decline. The first was to create and identify environmental refuges for the Growling Grass Frog, *Litoria raniformis*, which is listed as endangered in Victoria. Following extensive mapping of temperature and



Growling Grass Frog. Photo: Jenny Rolland

salinity of wetlands through central and northern Victoria, sites where chytrid would thrive were predicted. This was confirmed by the decreased incidence of the Growling Grass Frog detected at those wetlands. As a result, it was decided that the Winton Wetlands in northern Victoria with warmer temperature and higher salinity would provide a suitable refuge for this frog. So populations are being collected

from the Bendigo water treatment plant for quarantine and captive breeding before translocation to the Winton Wetlands, the "Task Frog Growler" project.

Another even more urgent project is aimed at protecting the critically endangered Corroboree Frog (*Pseudophryne corroboree*) by preventing transmission from host reservoir species. For this, fenced-off breeding areas are being built in the Kosciusko National Park where the frogs can build up numbers free from contact with fungal-infected but less susceptible species such as the Common Eastern Froglet (*Crinia signifera*). A similar strategy is being trialled for the Alpine Tree Frog (*Litoria verreauxii alpina*) which has the additional threat of habitat destruction from deer which trample and destroy water bodies.

Thirdly, Matt presented a project designed to improve frog survival by identifying factors that increase reproductive output. For this, the critically endangered Spotted Tree Frog (*Litoria spenceri*) was compared with the common, non-threatened Lesueur's Frog (*Litoria lesueuri*). Interestingly, elevation was shown to be an important influence. On balance, the effects of elevation on number of eggs/clutch, age at maturation and longevity,

meant that Lesueur's Frog was able to better survive at higher elevations. This study also revealed another important threat to frogs, predatory non-native fish! This has lead to a productive partnership with recreational fishing groups, reducing these fish species in selected areas, especially Brown Trout and Rainbow Trout. So, safe havens for our native frogs can be further defined by selection of areas free from non-native fish species, in addition to sites where factors improve reproductive success and where the chytrid fungus doesn't survive.

Finally in discussion, Matt addressed the question of "What can we do?" He urged us all to use the FrogID app to record frog calls wherever we are to better map the distribution of species and monitor their numbers. Cleaning our boots and spraying the soles with a fungicide (as we are already doing on our field trips for plant pathogens) is another way in which we can reduce the spread of the chytrid fungus. And of course we need to continue to nurture and protect our wonderful wetlands and the insects on which our frogs feed.

Jenny Rolland

# **Excursion: Remeasuring Smiths Reef Quadrats.**

Between 2001 and 2014 club stalwart, Ern Perkins established over a hundred permanently marked vegetation quadrats in forests across the region. His intention was to monitor and report on the impacts of fire on significant plant species and record the recovery and changes to the vegetation over time. In most cases there are pairs of plots, one inside and one outside proposed burn areas.

It was a cool overcast afternoon with rain threatening when about 25 members and friends, including a strong contingent from Maldon met at Smiths Reef dam, Maldon Historic Area, to examine and remeasure one of the Perkins vegetation quadrats. Among those present were Leslie Perkins, Richard Piesse and Denis Hurley, veterans from the original surveys who were able to provide valuable insights into how the quadrats were surveyed.

The location was chosen because there is a planned burn scheduled for this area and we already have three sets of prior data from before and after the last planned burn in 2009. The first challenge was relocating the corner marker. This is a short length of steel pipe that had been hammered into the ground near the base of a prominent tree to mark the north-west corner of the quadrat. Running lines were then laid out to delineate the boundary (20x20m) and then subdivide the quadrat into five strips 4m wide to make recording of the plants easier.

After laying out the quadrat some of the group carried out the count while the rest of the group explored the nearby area recording what was found in the wider area of the burn site.

During the quadrat survey we managed to find 33 species which included most of the species that had been found in the original surveys. Those species not recorded were herbaceous plants that were unlikely to be visible at this time of the year. We also found three species, *Xerochrysum viscosum*, *Luzula meridionalis* and *Pterostylis smaragdyna*, that were not on the original lists for the quadrat. Outside the quadrat more than thirty species of vascular plant, of which seven or eight were not found inside the quadrat, were recorded as well as mosses and numerous fungi and lichen. Those species recorded outside the quadrat included the rare *Eucalyptus aurifodina*, Small-leafed Brown Stringybark.

This excursion was a good introduction to the survey method and what is involved for the majority of those present who had little or no experience of formal botanical surveys. This will provide valuable information as to how our forests recover following fire and inform the ongoing debate about fuel reduction activities. We will write up the results of this survey for a future Castlemaine Naturalist and enter the data into the Victorian Biodiversity Atlas.

The club would like to have a group of keen plant enthusiasts who can repeat more of these surveys. Please contact the committee (castlemainefnc@hotmail.com.au) if you would like to help.

**Euan Moore** 





Left: Some of the group recording plant information during the survey. Right: Leslie Perkins setting up the running lines.

Observations
Cathrine Harboe-Ree McCay Res. Excursion - Fungi gallery



Opposite page, clockwise from top left: Graceful Parasol *Macrolepiota clelandii*, Elegant Blue Webcap *Cortinarius rotundisporous*, Trooping Crumble Cap *Coprinellus disseminatus*, *Mycena subgalericulata*, *Leucopaxillus eucalyptorum*, *Descolia* sp. (all at McCay except the Blue Webcap near Smiths Reef). Below left: Whirligig beetles at McCay Res.







Lou Citroen

Restless Flycatcher, July 4 (uncommon here)
Painted Button-Quail – one of two on May 18
near the Goldfields railway track.

**Russell Stanley** I went exploring the Fryerstown and Fryers Ridge areas on July 11 and found a selection of colourful fungi as well as a small number of Veined Helmet Orchid *Corybas diemenicus*. Unfortunately the millipedes had done all the flowers a fair bit of damage. Here are a couple of the less chewed specimens.





## **Kerrie Jennings**





Koala – seen a couple of weeks ago, Dunn's Track, Muckleford forest. Apparently seen occasionally in this area according to Geoff Park and Geoff Neville.

Right: White-bellied Cuckoo-shrike - dark phase, near Gough's Range.

# **Noel Young**

Rainbow Lorikeets – well they haven't taken over yet, but I often see a few around the Murphy Street trees in Wesley Hill, and sometimes in town. Here they are chewing seeds in a garden tree. I counted seven on this occasion. (July 30)





## **Mez Woodward**

Lepidoscia casemoth. They are so tiny, max 5mm to date, and typically feed on the lichen that is the same color as them, so devilishly hard to find.



### Binds of Sutton Grange July 2023

Migel Harland

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Leaders: Peter Former & Hosel Young.

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